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Toxicokinetics And Biotransformation Of Xenobiotics In Fishes

Toxicokinetics refers to the time-dependent processes and quantitative relationships that regulate how foreign substances are absorbed, distributed, metabolized, and eliminated in living organisms. In fish, these processes are fundamentally connected to their life in water and their specific physiological adaptations. Unlike land-dwelling creatures, fish are in constant interaction with dissolved and suspended foreign substances via their gill epithelium and digestive system. Furthermore, their metabolic activities take place at significantly lower body temperatures compared to terrestrial animals, resulting in unique detoxification timelines. The four essential stages of xenobiotic processing in fish—absorption, distribution, biotransformation, and excretion—collectively influence the biological outcome of pollutants and their ability to inflict damage. Each stage is influenced by chemical characteristics (such as lipophilicity, molecular weight, and ionization state), physiological aspects (including species-specific metabolism, organ functionality, and body composition), and environmental factors (like temperature, pH levels, dissolved oxygen content, and water hardness). In

contaminated aquatic environments, fish may be exposed to complex mixtures of various chemicals with differing structures and toxic effects, which can exceed their detoxification capacity and lead to unpredictable synergistic impacts that cannot be anticipated based on the effects of individual substances.

Absorption And Bioavailability

1. Primary Routes of Xenobiotic Entry

Fish take in xenobiotics primarily through two main avenues: absorption from the water via the gill epithelium and ingestion through the digestive system. The significance of these pathways varies greatly based on the lipophilicity of the chemicals and environmental factors. The gills serve as the main route for absorbing most waterborne organic substances, especially lipophilic materials. This prominence is due to both anatomical and physiological reasons—gills have a vast surface area (about 1000 times larger than the body's surface), exhibit high blood flow rates, and are closely linked to water through a relatively thin epithelial layer. Research involving lipophilic substances, with octanol-water partition coefficients ranging from 3.98 to 7.55, has shown that the rate at which gills absorb these chemicals is roughly five orders of magnitude higher than that of the gastrointestinal tract. Consequently, gills are expected to play a crucial role in determining the chemical load in the body for compounds that are moderately to highly lipophilic. The uptake of xenobiotics at the gill occurs through several mechanisms: passive transcellular diffusion influenced by concentration gradients, paracellular transport through tight junctions, and active transport through carrier-mediated systems. The significance of passive compared to active mechanisms is largely determined by the chemical characteristics of the substances and the availability of specific transporter proteins. Phase 0 transport, which involves active uptake through SLC carriers, and Phase III transport, which entails efflux through ABC transporters, work together with biotransformation enzymes to regulate intracellular levels and aid in the removal of substances. In cultures of primary gill cells from rainbow trout, evidence has shown both passive and carrier-mediated uptake processes, with transepithelial resistance measurements validating the effective properties of the epithelial barrier.

2. Bioavailability Modifiers

The bioavailability of chemicals in natural water systems is influenced by more than just their concentration in water. Factors such as suspended sediments, dissolved organic matter (including humic substances), and particulate organic carbon significantly impact how chemicals are speculated and how available they are for organisms to absorb. Lipophilic chemicals that are attached to suspended sediment with low organic carbon levels remain available for uptake through gills, as shown in mass

balance studies where the chemical burden in fish surpassed what would be expected from dissolved concentrations alone. On the other hand, higher levels of dissolved organic matter may hinder the bioavailability of certain compounds by competing for passive diffusion; however, this effect varies depending on concentration and the specific chemical involved.

3. Tissue-Blood Partition Coefficients

After being absorbed, xenobiotics spread throughout various tissue compartments, influenced by the specific lipid composition, protein binding ability, and membrane permeability of the tissues. The tissue-to-blood partition coefficients (P_{tb}) are crucial elements in toxicokinetic modeling, as they influence how much of a chemical accumulates in target organs. Recent computational methods that combine different modalities and incorporate both categorical features and molecular descriptors have improved the ability to predict P_{tb} values for numerous species, tissues, and classes of xenobiotics. However, it is still vital to validate these predictions empirically for particular combinations of compounds and tissues.

Phase I Biotransformation: Oxidation, Reduction, And Hydrolysis

1. Cytochrome P450 System and Mixed-Function Oxidases

The cytochrome P450 (CYP) superfamily is the key Phase I biotransformation system in fish, playing a crucial role in the oxidation of lipophilic substances and the addition of functional groups that aid in further metabolism. These hemoproteins are mainly found on the membranes of the endoplasmic reticulum in hepatocytes and gill epithelial cells, using molecular oxygen to insert hydroxyl groups or carry out oxidative deamination, which significantly enhances the polarity of compounds. Fish exhibit 14 primary CYP isoforms, with a focus on the CYP1A, CYP2, and CYP3 families that are involved in the metabolism of xenobiotics, while other CYP variants are involved in managing the production of endogenous steroids and eicosanoids. The CYP1A subfamily plays a crucial role in the metabolism of xenobiotics and in the monitoring of regulatory biometrics. The expression of CYP1A is modulated by the aryl hydrocarbon receptor (AhR), which is a transcription factor activated by ligands that can bind to polycyclic aromatic hydrocarbons (PAHs), dioxins, and similar compounds. When AhR is activated, it moves to the nucleus, pairs with ARNT, and begins the transcription of xenobiotic-responsive elements situated upstream of CYP1A and other detoxification genes. This induction process is swift and depends on the dosage, leading

to a 5 to 53-fold increase in CYP1A protein levels after exposure to certain PAHs. The effectiveness of the induced enzyme is commonly assessed through ethoxyresorufin-O-deethylase (EROD) assays, which evaluate the oxidative deethylation of ethoxyresorufin into the fluorescent compound resorufin.

2. Additional Phase I Enzymes

In addition to CYP-mediated oxidation, flavin-containing monooxygenases (FMOs) facilitate the oxidative transformation of xenobiotics that contain nitrogen and sulfur. These enzymes convert the substrates into N-oxides and sulfoxides, which are often more readily combined in Phase II reactions. NAD(P)H:quinone oxidoreductases (NQOs) reduce quinone and similar compounds, while esterases and amidases break down ester and amide groups, respectively, making hydroxyl and amino groups available for further conjugation. The distribution of Phase I enzymes within the bilayer of the endoplasmic reticulum is highly important, placing these enzymes at the lipid-water interface where lipophilic substrates tend to accumulate. In fish, the liver shows the highest enzymatic activity, but significant Phase I function is also present in the gills, kidneys, and intestinal tissue, indicating various potential entry points for xenobiotics and the first-pass metabolic processes.

Phase II Biotransformation: Conjugation And Synthetic Reactions

1. Glutathione S-Transferases

Glutathione S-transferases (GSTs) facilitate the key Phase II conjugation process by linking the tripeptide glutathione to phase I metabolites and parent compounds that have electrophilic sites. This conjugation significantly enhances the molecular hydrophilicity and leads to the formation of glutathione conjugates that are easily identified by transporter proteins and subsequently expelled through urine or bile. Fish have several GST isoforms, each showing different substrate affinities and specific expression patterns in various tissues. The activity of GST is a sensitive biochemical indicator of Phase II metabolic capability and exposure to oxidative stress, often increasing after contact with pesticides, heavy metals, or other foreign substances. The journey of glutathione conjugates does not end with simple urinary elimination. A series of enzymatic modifications occurs, including the removal of glutamic acid by γ -glutamyltransferase, the excision of glycine by dipeptidases, and the N-acetylation of the remaining cysteine residue, resulting in mercapturic acid metabolites, which are the final excretory form of many xenobiotics. Recent studies have discovered mercapturic acid pathway

products in fish exposed to complex petroleum hydrocarbons and specific polycyclic aromatic hydrocarbons (PAHs), highlighting the importance of this pathway in aquatic organisms that had previously been undervalued.

2. UDP-Glucuronosyltransferases and Other Conjugation Systems

UDP-glucuronosyltransferases (UGTs) facilitate Phase II glucuronidation by adding glucuronic acid units, obtained from UDP-glucuronic acid, to xenobiotic metabolites that feature hydroxyl, carboxyl, or amino groups. This process significantly raises the molecular weight and greatly improves water solubility. Various UGT isoforms are expressed in fish with overlapping substrate specificities, which aids in the biotransformation of a wide range of pharmaceutical residues, phenolic substances, and phase I metabolites. Recent untargeted metabolomic techniques have allowed for thorough profiling of fish glucuronidation products, uncovering a diversity of biotransformation pathways that were previously difficult to access with targeted analytical techniques. Sulfotransferases promote the sulfonation of phenolic and other nucleophilic substrates, leading to the formation of sulfate esters, which exhibit remarkably increased water solubility. N-acetyltransferases facilitate the N-acetylation of amines and nitrogen atoms within amides. Methyl transferases play a role in the phase II metabolism of specific compounds through methylation processes. The significance of these various Phase II pathways differs depending on the structure of the xenobiotic, its concentration, and the specific expression patterns of enzymes in different tissues.

Organ-Specific Biotransformation Capacity

The multidrug resistance-associated proteins (MRPs) and P-glycoproteins are part of the ATP-binding cassette (ABC) transporter superfamily. They play a vital role in actively transporting phase II metabolites and unaltered xenobiotics from the hepatocyte membrane into bile. These transporters, which rely on energy, link the process of chemical transport to ATP hydrolysis, allowing substrates to move against their concentration gradients. ABC transporters are essential in the removal of xenobiotics by decreasing the intracellular levels of potentially harmful substances and metabolites, thereby preventing their accumulation in hepatocytes and other vulnerable tissues. In fish, the expression of ABC transporters can be triggered by exposure to xenobiotics and shows patterns that vary by tissue type. Both hepatic and extra-hepatic ABC proteins are involved in defending against xenobiotics, playing especially significant roles in the intestinal epithelium, where they regulate absorption, and in the gill

epithelium, where they inhibit the internal buildup of chemicals from water. Importantly, numerous environmental pollutants act as inhibitors of ABC transporter function, including some pesticides, heavy metals, and pharmaceutical items, which can hinder the detoxification ability and increase the risk of bioaccumulation of other co-occurring chemicals.

Hepatic Metabolism As Primary Site

The liver is the main organ responsible for the biotransformation of xenobiotics in fish, similar to other vertebrates. This is attributed to its anatomical location, which allows it to receive portal blood from the gastrointestinal tract, as well as its high concentration of Phase I and Phase II enzymes. The activity of hepatic phase I enzymes, such as EROD and CYP-dependent monooxygenase activity, typically outstrips that of extra-hepatic sites by significant margins. Hepatocytes grown in a three-dimensional spheroid setup exhibit better expression and functionality of biotransformation enzymes than those in standard monolayer cultures. This configuration preserves metabolic capabilities for longer durations and facilitates more physiologically relevant toxicity evaluations.

- **Extra-Hepatic Biotransformation Sites**

The liver plays a central role in overall metabolic capability; however, tissues outside the liver also significantly aid in the biotransformation of xenobiotics, especially for substances that enter the body through non-portal pathways. The gill epithelium serves as the main entry point for waterborne chemicals, showing considerable Phase I and Phase II metabolic activity. Cultures of primary gill cells and immortalized gill cell lines, such as ASG-10 from Atlantic salmon, are capable of metabolizing xenobiotics, including polycyclic aromatic hydrocarbons (PAHs), and exhibit biotransformation abilities similar to certain liver preparations. The epithelial cells of the gastrointestinal tract also express Phase I and Phase II enzymes, playing a vital role in the first-pass metabolism of dietary xenobiotics. Intestinal ATP-binding cassette (ABC) transporters are crucial for controlling the bioavailability of dietary chemicals through active efflux, which prevents the full absorption of ingested toxins. The kidneys engage in the active reabsorption and metabolism of specific xenobiotics, while less thoroughly researched tissues, such as the brain, heart, and endocrine glands, demonstrate a capacity for biotransformation that aligns with their risk of xenobiotic exposure and metabolic requirements.

Xenobiotic-Specific Biotransformation Pathways

1. Pesticides and Organophosphate Compounds

Pesticide pollution leads to ongoing exposure in agricultural watersheds, with numerous chemical types currently in use. The transformation of pesticides occurs through various pathways influenced by their chemical makeup; nonetheless, oxidative transformation involving CYP-mediated processes serves as a frequent initial phase for many pesticide categories. Organophosphate pesticides are subject to hydrolysis and oxidative transformation, with significant toxic impacts resulting from the inhibition of acetylcholinesterase (AChE), the enzyme tasked with breaking down acetylcholine at neuromuscular junctions. Interestingly, certain organophosphates are converted into more toxic oxon metabolites through CYP-mediated oxidation, enhancing their toxic potential instead of diminishing it. This conversion leads to variations in pesticide toxicity that depend on species and individual metabolic capabilities. The toxicity caused by pesticides encompasses various mechanisms that exceed mere direct enzyme inhibition. The production of reactive oxygen species (ROS) from mitochondrial disruption and the formation of phase I metabolites results in oxidative stress, which overloads antioxidant defenses and triggers lipid peroxidation, protein damage, and DNA impairment. The reduction of Na^+/K^+ -ATPase activity interrupts osmotic stability, while immunosuppression diminishes resistance to diseases. Additionally, changes at the transcriptional level affect the expression of detoxifying enzymes, antioxidants, and genes related to the immune response.

2. Heavy Metals and Metalloids

In sharp contrast to organic xenobiotics that undergo enzymatic biotransformation via Phase I-II-III processes, heavy metals such as cadmium, lead, copper, zinc, and mercury cannot be chemically converted into less harmful forms. Instead, fish manage metal toxicity by producing metallothionein, changing how their organs absorb and distribute metals, and excreting them through feces or urine. The physiological processes that contribute to metal toxicity—such as binding to sulfhydryl groups on critical enzymes, displacing necessary cofactors, and generating reactive oxygen species through Fenton chemistry and enzyme inhibition—are clearly different from xenobiotic biotransformation pathways. Heavy metals accumulate in fish tissues through mechanisms like gill uptake from water, skin absorption, and dietary intake, with the extent of each method varying based on metal type, water chemistry, and feeding patterns. Notable species-specific variations in metal accumulation exist; demersal bottom-feeding fish exposed to contaminated sediments tend to accumulate metals more significantly

than pelagic species, whereas high-metabolism predatory fish may show higher metal concentrations even with less direct exposure to the environment. Patterns of tissue distribution indicate that numerous metals tend to accumulate in the liver, with notable increases also observed in the gills, kidneys, and bones. The bioaccumulation factors (BAF), which compare tissue concentration to environmental concentration, differ significantly among species, showing values that range from 100 to over 100,000 for certain metals in specific species. The process of biomagnification of metals via trophic transfer within the food web heightens the toxicological threat to top predators and humans who eat contaminated seafood.

Biomarkers Of Xenobiotic Exposure And Effect

Advanced understanding of toxicokinetics has facilitated the creation of physiologically-based multicompartment models that simulate the uptake, distribution, metabolism, and elimination of foreign substances in fish. These models combine various factors, including chemical characteristics (such as K_{ow} , molecular weight, and rates of chemical transformation), physiological details (like organ sizes, blood flow rates, and partition coefficients), and environmental variables to forecast tissue concentrations over time. PBTK models for fish show greater predictive capability compared to simpler single-compartment models, especially in non-steady-state scenarios and for substances with moderate to high lipophilicity. By utilizing tissue-specific partition coefficients based on lipid content, rather than relying solely on simple K_{ow} correlations, the precision of predictions across multiple tissues is improved. Recent studies have applied PBTK modeling to antifungal pesticides like imazalil and prochloraz, as well as bisphenols, making it possible to predict internal tissue dosages significant for assessing mechanistic toxicity. However, uncertainties in the models are considerable, particularly for fish species that have not been thoroughly characterized physiologically, for xenobiotics that exhibit metabolic interactions, and for complicated environmental mixtures with numerous chemicals. Despite these challenges, PBTK frameworks serve as valuable tools for merging mechanistic toxicological insights and aiding in risk assessment processes.

Phase I Enzyme Induction Biomarkers

Hepatic ethoxyresorufin-O-deethylase (EROD) activity serves as the most commonly used indicator of polycyclic aromatic hydrocarbon (PAH) and other aryl hydrocarbon receptor (AhR)-active compound exposure in fish populations. This enzyme facilitates the oxidative deethylation of ethoxyresorufin, producing the fluorescent compound resorufin, which can be

quantified using spectrofluorometry with sufficient sensitivity to identify the effects of contaminants at environmentally relevant exposure levels. In fish collected from polluted locations, EROD activity is found to be elevated 36 to 53 times higher than in reference controls, allowing for a quantitative evaluation of PAH bioavailability and biological activity. The effectiveness of EROD as a biomarker for exposure relies on its responsiveness to AhR-active substances (such as PAHs, dioxins, and PCBs), its inherent expression at detectable baseline levels, and its consistent presence across various fish species. Nevertheless, EROD activity indicates recent exposure over periods ranging from weeks to months, and the enzyme's activity tends to revert to baseline levels fairly quickly after contaminants are removed. Furthermore, multiple classes of xenobiotics can stimulate CYP1A expression via various signaling pathways, which may affect its specificity.

CYP3A activity, often evaluated through the hydroxylation of testosterone or other CYP3A-specific compounds, acts as an alternative phase I biomarker with wider substrate specificity and susceptibility to induction by various classes of xenobiotics, including medications and certain pesticides. Additionally, gill cell-based biomarkers for phase I enzyme activity offer tissue-specific insights into the potential internal accumulation of chemicals in water.

Phase II Enzyme Activity and Oxidative Stress Biomarkers

Glutathione S-transferase (GST) activity, observed in liver and gill tissues, increases after exposure to various groups of xenobiotics and acts as a sensitive measure of phase II metabolic activation and oxidative stress. In contrast to phase I enzymes, which exhibit selective induction based on the type of xenobiotic, GST activity tends to increase broadly in response to oxidative challenges, biomarkers of oxidative stress, and chemical exposures that disrupt the cellular redox balance. Malondialdehyde (MDA), a byproduct of lipid peroxidation, builds up in fish tissues during exposure to oxidative stress and serves as a quantitative measure of damage to cellular lipids. Reduced glutathione (GSH), which is the nucleophilic substrate for GST reactions, gets depleted during detoxification processes that heavily utilize antioxidants, and lower levels of hepatic GSH indicate metabolic impairment. When combined, GST activity and the ratios of MDA to GSH offer a comprehensive evaluation of phase II metabolic capacity and the status of oxidative stress. Additionally, acetylcholinesterase (AChE) activity in red blood cells or nervous tissue declines following exposure to organophosphate and carbamate pesticides, indicative of the irreversible phosphorylation of the enzyme's active site's catalytic serine residue. AChE inhibition exceeding 20-30% below baseline levels serves as significant

evidence of neurotoxic pesticide exposure and disruption of neuromuscular transmission functionality.

1. Genotoxic Biomarkers

DNA adducts, which are covalent alterations of DNA bases caused by reactive xenobiotic metabolites, build up in the liver cells of fish that have been exposed to genotoxic substances like PAHs and pesticides. The measurement of these adducts using ³²P-postlabeling or immunochemical techniques offers direct proof of contact with DNA-reactive agents. While DNA adducts serve as sensitive early indicators of genotoxic risk, their longevity is influenced by the reactivity of the chemicals and the effectiveness of DNA repair processes, necessitating careful consideration over time. Micronuclei (MN) are fragments of extranuclear chromatin that arise from breaks in DNA strands or chromosomal abnormalities triggered by exposure to genotoxic agents. Counting micronuclei in fish blood cells, such as erythrocytes, provides a quantitative measure of chromosomal damage and mutagenic potential. The frequency of sister chromatid exchanges (SCE), assessed through cytogenetic studies, reflects the processes of homologous recombination and DNA strand break repair; an increase in these exchanges suggests unrepaired DNA damage.

Xenobiotic Mixture Toxicity And Synergistic Effects

1. Non-Additive Mixture Toxicity

Fish in natural water environments typically do not encounter isolated xenobiotics; instead, they face intricate blends of chemicals, such as polycyclic aromatic hydrocarbons (PAHs), pesticides, heavy metals, persistent organic pollutants, and pharmaceutical residues. The toxicity of these chemical combinations often diverges significantly from what would be predicted based on additive models, with both synergistic (where toxicity exceeds the sum of individual effects) and antagonistic (where toxicity is less than the sum) interactions being quite common. Mixtures of polycyclic aromatic hydrocarbons illustrate the complexities of toxicology regarding mixtures. When fish embryos and larvae are exposed to straightforward binary mixtures of PAHs—such as an AhR agonist (benzo[a]pyrene; BaP) and another PAH (fluoranthene)—the resulting cardiotoxicity is markedly greater than that caused by BaP on its own, even though fluoranthene has minimal effects when tested separately. This enhanced toxicity is due to fluoranthene's ability to inhibit CYP1A activity, which hinders the biotransformation and elimination of BaP while still permitting the activation of AhR by BaP

to occur without restriction. On the other hand, when Atlantic haddock are exposed to a combination of phenanthrene and the CYP1A inducer 3-methylchrysene, there is a fivefold increase in the levels of phenanthrene metabolites. This combination consistently results in more significant morphological abnormalities and cardiotoxicity compared to exposure to phenanthrene alone. This example illustrates how bioaccumulation can occur through metabolic inhibition, where one substance inhibits the elimination of another, functioning as a key mechanism that leads to synergistic effects in mixtures.

2. Metabolic Interactions and Enzyme Inhibition

Pharmaceutical substances that act as CYP inhibitors, such as specific azole antifungals like ketoconazole and clotrimazole, as well as prochloraz, can significantly impact the toxicokinetics of concurrently exposed xenobiotics by diminishing their liver clearance and extending their biological half-life. Predictive mathematical toxicokinetic models that include enzyme inhibition have effectively forecasted the delayed removal of the aromatic hydrocarbon β -naphthoflavone (BNF) in fish liver cells that are simultaneously exposed to the CYP inhibitor nocodazole (NOC). This illustrates a prolonged interaction with cytochrome P450 active sites and a decrease in BNF clearance. The important toxicological consequence is considerable: exposure in the environment to mixtures of metabolic enzyme inducers and inhibitors can enhance the toxicity of individual elements through competitive inhibition and modified clearance dynamics. This mechanism is often overlooked in environmental risk evaluations, which usually presume that the presence of xenobiotics does not affect each other.

Ecosystem-Level Consequences Of Altered Xenobiotic Biotransformation

1. Population-Level Effects Through Reproductive Disruption

The ability of individual fish to metabolize contaminants plays a crucial role in determining toxicological risk; however, the broader impact on fish populations can be significant, leading to disruptions in reproductive systems and developmental anomalies. Endocrine-disrupting chemicals (EDCs), while having limited effects on the survival of individual fish, can severely interfere with reproductive growth and the processes of sexual differentiation. Some EDCs even cause complete sex reversal in males, promoting feminization at concentrations typically found in the environment. The hypothalamic-pituitary-gonadal (HPG) axis, which is the primary neuroendocrine system that governs reproduction in vertebrates, is particularly vulnerable to the effects of EDCs. Changes in the expression of GnRH (gonadotropin-releasing

hormone), the responsiveness of the pituitary gland, and steroid production work together to hinder gamete formation, diminish fertilization rates, and lower the survival rates of offspring. Consequently, even when there are slight impacts on adult fish survival, these factors can lead to a significant decline in reproductive success at the population level. Furthermore, disruptions in thyroid function caused by pesticides and other foreign substances worsen reproductive issues, as thyroid hormones are essential for regulating growth, development, and metabolic balance. The combination of direct effects on reproduction and thyroid-related developmental disruptions creates an increased risk for fish populations.

2. Community Structure and Food Web Dynamics

The accumulation of heavy metals and persistent organic pollutants escalates systematically across different trophic levels due to biomagnification within food webs. Apex predatory fish, which are at the top of these trophic levels, show contaminant body burdens that reflect both direct exposure to environmental pollutants and bioaccumulation from all their prey. As a result, predatory fish species exhibit higher levels of contaminants and greater health issues compared to species from lower trophic levels in the same habitats. This variation in vulnerability leads to changes in community structure: sensitive species decline in polluted environments, while more adaptable generalists become more prevalent. The outcome is a simplification of communities with diminished species diversity, weakened ecosystem resilience, and modified trophic interactions. The decline of apex predators due to contaminant accumulation can initiate trophic cascades, resulting in an increase of mesopredators and herbivorous fish, and a decrease in primary productivity due to changed grazing behaviors.

3. Disease Susceptibility and Population Viability

Immunosuppression caused by xenobiotics—driven by various mechanisms such as oxidative stress, hormonal disruption, and direct impacts on lymphoid tissues—diminishes the disease resistance of fish populations that are exposed. Field studies of contaminated fish show higher rates of parasitism, a greater prevalence of infectious diseases, and a weakened response to environmental pressures when compared to reference populations. This impairment in their immune system raises the possibility of disease outbreaks at the population level that would not typically occur in unexposed groups. The interplay of diminished reproductive rates, increased mortality in juveniles, higher disease-related deaths in adults, and the loss of habitat collectively endangers the sustainability of these populations. Mathematical models that take into account the effects of contaminant exposure suggest that population collapse can happen

at contamination levels lower than those causing noticeable mortality, emphasizing the subtle yet harmful impact of chronic, sublethal xenobiotic exposure on population health.

Environmental Monitoring And Assessment Integration

1. Multi-Biomarker Approaches

Modern environmental evaluations increasingly incorporate various biomarkers across different biological levels—from molecular (such as gene expression and enzyme activity) to cellular (like DNA damage and histopathology) and organismal (including growth, behavior, and reproduction)—instead of depending on just one indicator. A thorough biomarker strategy that simultaneously examines EROD activity (indicating phase I exposure), GST activity and MDA levels (reflecting phase II capacity and oxidative stress), plasma VTG and GSI (signifying reproductive disruption), along with genotoxic endpoints (such as DNA adducts and micronuclei), offers a well-rounded assessment of the hazards, exposure, and effects of contaminants. This multi-biomarker method improves biological relevance because no single biomarker fully encompasses the intricate nature of toxicity induced by xenobiotics. The integration of various biomarkers in riverine environments has effectively differentiated between unpolluted and contaminated sites, as well as monitored the progress of pollution remediation efforts.

2. Field Populations as Sentinels

Wild fish populations in polluted water bodies exhibit a range of biomarker changes that indicate exposure to environmental chemicals. In the Paraíba do Sul River, downstream from industrial discharge points, wild fish displayed a 53-fold increase in hepatic EROD activity when compared to reference locations, suggesting significant exposure to PAHs and other AhR-ligands. At the same time, fish condition factors (body weight in relation to length) showed a decrease, and gonadosomatic indices dropped in the most polluted areas, offering evidence of the effects of pollution at the organism level. Utilizing fish as indicators of environmental health is a cost-effective and biologically meaningful method of monitoring, surpassing traditional chemical analyses. Fish naturally accumulate exposure over time, preferentially gather bioavailable chemical components, and react to combined effects that typical water column assessments may overlook. Consequently, fish-based evaluations of environmental quality have become essential to regulatory policies around the world.

3. Predictive Toxicology Integration

New developments in PBPK modeling, computational toxicology, and adverse outcome pathway (AOP) frameworks offer the potential to incorporate insights into biotransformation into quantitative hazard and risk assessment. As the details of biotransformation processes become clearer through proteomics, metabolomics, and functional genomics techniques, the ability to predict xenobiotic-specific impacts will improve regulatory choices and methods for ecosystem conservation.

Conclusions And Future Perspectives

Grasping the toxicokinetics and biotransformation processes of xenobiotics in fish is essential for evaluating contamination risks to aquatic ecosystems and predicting the broader impacts of chemical pollution on fish populations. Fish biotransformation systems have developed to manage natural plant toxins and internal metabolic substances; however, in today's aquatic environments, they confront significant and unprecedented exposures to a multitude of synthetic chemicals with unique structures and mechanisms. The dominant belief that biotransformation is always a detoxifying process requires considerable reevaluation, particularly when metabolic activation leads to more harmful metabolites than the original compounds, when interactions among mixtures surpass detoxification abilities, and when endocrine disruptors have impacts that are unrelated to traditional biotransformation pathways. New findings highlighting the role of microbiota in xenobiotic biotransformation, intricate synergistic effects of mixtures, and unanticipated metabolic processes (such as the mercapturic acid pathway and the production of polycyclic aromatic acids) not only enhance our mechanistic understanding but also expose significant gaps in knowledge. Future research priorities include systematic evaluation of biotransformation capacity across diverse commercially important fish species, development of *in vitro* systems better predicting *in vivo* toxicokinetics, integration of biotransformation understanding into population genetic models predicting evolution of pollutant tolerance, and expanded investigation of realistic chemical mixture toxicity. The critical importance of fish biotransformation capacity to population viability and ecosystem integrity makes this fundamental physiological process essential to aquatic conservation and the protection of freshwater and marine ecosystems

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