

## Journal of Science Innovations and Nature of Earth

Journal homepage: www.jsiane.com

# TOXIC EFFECT OF BIFENTHRIN INSECTICIDES ON THE LIVER ENZYMES IN CHANNA PUNCTATUS (BLOCH.)

#### Prem Sagar<sup>1</sup>, Shalini Yadav<sup>1</sup>, Shekhar Biswas<sup>2</sup>, Neelam<sup>3</sup> and Anand Pratap Singh<sup>2</sup>

<sup>1</sup>Department of Zoology, Government PG College, Jalesar, Etah Affiliated to RMPSU Aligrah, Uttar Pradesh, India <sup>2</sup>Department of Zoology, Agra College, Agra, Affiliated to Dr. Bhimrao Ambedkar University, Agra, Uttar Pradesh, India <sup>3</sup>Department of Zoology, N.R.E.C. College, Khurja (Bulandshahr), Affiliated to C.C.S. University, Meerut, UP. India. Email: premlovesagar@gmail.com

www.doi.org/10.59436/https://jsiane.com/archives3/3/94

#### **Abstract**

Pollution levels in today's world are rapidly rising, endangering not just the natural world but also human and animal health. Dangerous contaminants include things like metals, human waste, and agricultural runoff with pesticides. Treatment of Channa punctatus (Bloch.) with bifenthrin insecticide for 7, 15, 30, 45, and 60 days resulted in significant increases in the levels of Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), and Alkaline Phosphatase (ALP), as compared to the control group.

Keywords: Channa punctatus, bifenthrin, Alanine Aminotransferase, Aspartate Aminotransferase, Alkaline Phosphatase

Received 05.04.2023 Revised 20.07.2023 Accepted 23.09.2023

#### Introduction

The amount of pollution in the modern world is rising quickly, which is dangerous for both human and animal health as well as the environment. Heavy metals, residential sewage, and agricultural runoff including pesticides are among the pollutants deemed harmful (Sagar et al., 2017). Both freshwater and marine aquatic habitats are affected by these contaminants. Pesticides make up a sizable component of these. In light of the world's steadily growing population, it is vital to solve problems related to food scarcity. One of the main Sustainable Development Goals (SDGs) and Millennium Development Goals (MDGs) is to end hunger. Pesticides are one of the many methods and chemicals used today to increase agricultural production. They are used to prevent, kill, and repel pests that harm crops, lower yields, and degrade food quality (Ullah et al., 2019). Pesticides from agricultural areas cause the water quality to decline, endangering the health of aquatic species and biodiversity. Although the types and quantities of these pesticides are always changing, their use is steadily rising worldwide. Different classes of these pesticides, such as insecticides, fungicides, herbicides, nematicides, rodenticides, acaricides, etc., are used against various target populations. In terms of employability, however, pesticide use outperforms the other classes (Ullah et al., 2018). There are several types of insecticides used, including synthetic pyrethroids and organophosphates. Because organophosphates have severe neurotoxic effects on mammals, their use was gradually phased out. As a result, the use of synthetic pyrethroids increased (Werner and Moran, 2008). According to the report's findings (van den Berg et al., 2012), pyrethroid use increased with time, reaching more than  $1500\ MT$  in  $2009\ from\ 100\ MT$  in 2000.

In the 1970s, synthetic pyrethroids (an artificial version of the decorative plant Chrysanthemum cinerariaefolium) were created. They were initially used for agricultural purposes, such as to protect food, but over time, they were also used for a variety of non-agricultural purposes, such as controlling animal ectoparasites, lice shampoos, mosquito repellents, and insect control sprays (Velisek et al., 2009; Brander et al., 2016a; Brander et al., 2016b). Due of their little hazardous effects on birds and mammals, the usage of synthetic pyrethroids is continuously increasing. These insecticides are used the second-highest (Wang et al., 2017). Numerous investigations assessed and verified the occurrence of synthetic pyrethroids in various biospheres. Some research (Ueyama et al., 2009; Corcellas et al., 2012; Wielgomas and Piskunowicz, 2013) and aquatic organisms have proven their presence in human breast milk and urea.

According to Wang *et al.* (2017), bifenthrin is a moderately dangerous (class II) chemical that the WHO has approved for usage in the general population. One of the main sources of food and nutrients for humans is fish. The nutritional value of various fishes is influenced by their biochemical makeup, which includes things like proteins, carbs, vitamins, and minerals (Sagar *et al.*, 2018). The *Channa punctatus* (Bloch) is chosen for the current investigation due to its simplicity in handling and availability. When travelling through a river that receives trash from nearby human settlements and industry, the fish "*Channa punctatus* (Bloch)" are the most vulnerable of all aquatic creatures to such pollution. The fish's tissues are

easily damaged by water contaminants. By disrupting its neurological system, the pyrethroid pesticide bifenthrin is mostly used to combat red imported fire ants. It is extremely poisonous to aquatic life. It is permitted to be sold for everyday use even though it is categorized as a restricted use chemical in the US, provided that the product has low bifenthrin content.

#### **Material and Methods**

Bifenthrin pesticide will be employed for the current study. From ponds near Agra, live Channa punctatus specimens were moved. The experiment used fish that were almost the same size and weight to represent a similar age group as a constant factor and to examine the effects of different pesticide treatments. To get rid of any potential skin infections, fish were cleansed in a solution containing 0.1% KMnO<sub>4</sub>. They were then applied to aquariums that held water after being rinsed with ordinary water. The fish were split into two groups: control group and treatment group. The control group and treated group were then separated into five smaller groups, each with six fish. A total of 6 fish make up the control group (A), whereas 30 fish make up the treated group, divided into 5 subgroups (B, C, D, E, and F), each with 6 fish. On days 7, 15, 30, 45, and 60, respectively, the treated and control groups' rats will be dissected out. After cutting the caudal peduncle of living fish with scissors, blood samples were taken in order to separate the serum for the testing of liver enzymes among all the groups. Liver enzymes; Alkaline Phosphatase (ALP) was estimated by the method of King and King's (1954) while the estimation of Alanine Aminotransferase (ALT) and Aspartate Aminotransferase (AST) was done by the method of Reitman and Frankel's (1957).

#### Results

According to the current study, fish treated with bifenthrin have elevated liver enzyme levels when compared to fish in the control group. Alkaline phosphatase (ALP), Alanine Aminotransferase (ALT), and Aspartate Aminotransferase (AST) levels significantly and highly significantly increased after 7, 15, 30, 45, and 60 days, respectively, according to the current data.

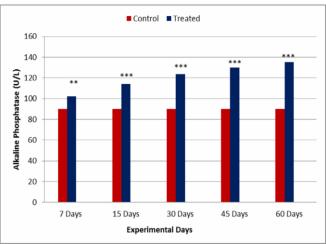
**Table I :** Effect of Bifenthrin on Alkaline Phosphatase (U/L) in *Channa punctaus* (Bloch) after 7, 15, 30, 45 and 60 days Exposure

S.	No. of days	No. of rats	Range Mean ± S.Em.	
No.			Control	Treated
1	7 Days	10	87.64–92.89 (90.07 ± 0.48)	99.87–104.81 (102.02 ± 0.48) ↑**
2	15 Days	10	87.64–92.89 (90.07 ± 0.48)	110.24 - 116.87 $(114.11 \pm 0.58) \uparrow ***$
3	30 Days	10	87.64–92.89 (90.07 ± 0.48)	119.44 - 126.81 (123.62 ± 0.62) $\uparrow$ ***
4	45 Days	10	87.64–92.89 (90.07 ± 0.48)	127.63 - 132.44 $(130.09 \pm 0.48) \uparrow ***$
5	60 Days	10	87.64–92.89 (90.07 ± 0.48)	133.01 - 139.45 $(135.45 \pm 0.66) \uparrow ***$

S.Em. = Standard Error of Mean

↑ Increase \*\* Significant

\*\*\* Highly Significant



**Fig. 1 :** Effect of Bifenthrin on Alkaline Phosphatase (U/L) in *Channa punctaus* (Bloch) after 7, 15, 30, 45 and 60 days Exposure

**Table II:** Effect of Bifenthrin on Alanine Aminotransferase (U/L) in *Channa punctaus* (Bloch) after 7, 15, 30, 45 and 60 days Exposure

S. No.	No. of Days	No. of Rats	Range Mean ± S.Em.	
			Control	Treated
1	7 Days	10	21.84–33.81 (27.04 ± 1.15)	38.78 - 43.11 $(40.67 \pm 0.48) \uparrow **$
2	15 Days	10	21.84–33.81 (27.04 ± 1.15)	42.59 – 47.51 (44.66 ± 0.47) ↑***
3	30 Days	10	21.84–33.81 (27.04 ± 1.15)	47.24 – 52.05 (49.68 ± 0.51) ↑***
4	45 Days	10	21.84–33.81 (27.04 ± 1.15)	50.29 - 54.93 (52.80 ± 0.45) ↑***
5	60 Days	10	21.84–33.81 (27.04 ± 1.15)	53.44 - 56.49 (54.86 ± 0.31) ↑***

S.Em. = Standard Error of Mean

↑ Increase \*\* Significant

\*\*\* Highly Significant

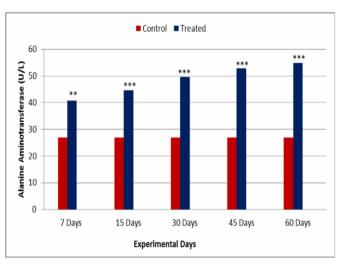


Fig. 2: Effect of Bifenthrin on Alanine Aminotransferase (U/L) in *Channa punctaus* (Bloch) after 7, 15, 30, 45 and 60 days Exposure

**Table III :** Effect of Bifenthrin on Aspartate Aminotransferase (U/L) in *Channa punctaus* (Bloch) after 7, 15, 30, 45 and 60 days Exposure

S. No.	No. of Days	No. of Rats	Range Mean ± S.Em.	
			Control	Treated
1	7 Days	10	22.41 - 26.56	35.87 – 39.81
1			$(24.61 \pm 0.46)$	$(37.97 \pm 0.43) \uparrow **$
2	15 Days	10	22.41 - 26.56	39.53 - 47.02
			$(24.61 \pm 0.46)$	$(43.93 \pm 0.79) \uparrow ***$
3	30 Days	10	22.41 - 26.56	48.63 - 52.33
			$(24.61 \pm 0.46)$	$(50.68 \pm 0.40) \uparrow ***$
4	45 Days	10	22.41 - 26.56	52.69 - 58.29
			$(24.61 \pm 0.46)$	$(54.49 \pm 0.56) \uparrow ***$
5	60 Days	10	22.41 - 26.56	55.51 - 61.02
			$(24.61 \pm 0.46)$	$(58.27 \pm 0.54) \uparrow ***$

S.Em. = Standard Error of Mean

↑ Increase \*\* Significant \*\*\* Highl

t \*\*\* Highly Significant

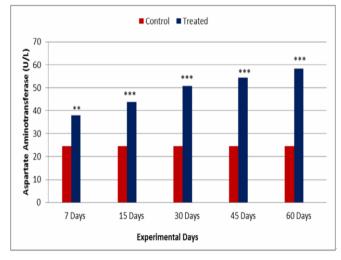


Fig. 3: Effect of Bifenthrin on Aspartate Aminotransferase (U/L) in *Channa punctaus* (Bloch) after 7, 15, 30, 45 and 60 days Exposure

#### **Discussion**

The liver, the body's largest gland, is crucial to metabolism, bile production, blood purification, yolk synthesis, detoxification, glycogenolysis, gluconeogenesis, deamination, and urea formation, among other processes. Any damage to the liver will impede all of these processes. Alkaline phosphatase (ALP), Alanine Aminotransferase (ALT), and Aspartate Aminotransferase (AST) levels in the liver have increased significantly in the current investigation; these enzymes are frequently linked to liver dysfunction or injury. An important part of the metabolism of non-essential amino acids is played by aminotransferases (ALT & AST). The cytosolic enzyme ALT, which is found in hepatocytes, is involved in intracellular metabolism. According to Nafiu et al. (2020), AST is primarily found in the liver, skeletal muscles, RBCs, and is associated with mitochondrial damage. ALP is an enzyme that is membrane-bound and may operate as both a hydrolase and a transphosphorylase. It is present on all cell membranes, where active transport takes place. The liver, biliary tract epithelium, bone, and intestinal mucosa all contain high levels of ALP. Through hepatic cell proliferation, serum ALP activity rises in cases of bile duct obstruction and hepatic cell injury (Ahmad et al., 2011). It is mostly found in the liver and is essential for the metabolism and production of macromolecules.

Consistent with the current findings At experimental times, diazinon-treated groups had significantly greater aspartate aminotransferase and alanine aminotransferase activity and glucose levels compared to the control group, as reported by Banaee et al., 2011. Further evidence that phyruvalate creation contributes more than oxaloacetate formation comes from the observation that SGPT activity was considerably higher than SGOT by Ghanim et al., 2020. After four weeks of exposure to fenvalerate. Zebra fish showed altered acid phosphatase and alkaline phosphatase activity in the liver and gills. Furthermore, the results presented here are supported by Bálint et al., 1995. The AChE activity of the blood serum decreased by 20% after 3 days of in vivo deltamethrin therapy, whereas the GOT and LDH activities increased by 2.5-fold (72-h sample) and 1.5fold (6-h sample), respectively, in comparison with the control carp. Six-hour blood samples showed 30% higher glucose levels compared to controls. The measurements showed that the fish were fully recovered after only 24-48 hours in clean water.

Glycogen metabolism also involves alkaline phosphatase in the liver. Glycogen production is stimulated while phosphorylase enzymes are inhibited by this enzyme. Under stress, the liver's alkaline phosphatase activity decreases to satisfy the body's increased energy needs, or the rate of transphosphorylation slows down, or the body's oxidative phosphorylation becomes uncoupled. Alkaline phosphatase activity in the liver of H. fossilis was shown to be diminished following exposure to 0.3-0.5 g L1 cypermethrin, as reported by Saha and Kaviraj (2009). Brain tissue from cypermethrin-exposed Labeo rohita showed decreased alkaline phosphatase activity, as was similarly documented by Das and Mukherjee (2003). However, it has been shown that fish exposed to cypermethrin show an increase in alkaline phosphatase activity (Frat et al., 2011; Loteste et al., 2013). In addition, Channa punctatus treated to cypermethrin and -cyhalothrin showed increased activities of lactate dehydrogenase (LDH), acid and alkaline phosphatases (Kumar et al., 2012). An elevation of alkaline phosphatase in the blood has been linked to liver necrosis and the subsequent release of the enzyme into the bloodstream (El-Sayed et al., 2008).

### References

Ahmad, L., Khan, A. and Khan, M.Z. (2011). Cypermethrin Induced Biochemical and Hepato-Renal Pathological Changes in Rabbits. *Int. J. Agric. Biol.*, 13(6): 865-872.

Al-Ghanim, K.A., Shahid Mahboob, P. Vijayaraghavan, F.A. Al-Misned, Young Ock Kim, Hak-Jae Kim (2020). Sub-lethal effect of synthetic pyrethroid pesticide on metabolic enzymes and protein profile of non-target Zebra fish, Danio rerio, Saudi Journal of Biological Sciences, 27(1): 441-447.

Bálint, T., Szegletes, T., Szegletes, Zs., Halasy, K. and Nemcsók, J. (1995). Biochemical and subcellular changes in carp exposed to the organophosphorus methidathion and the pyrethroid deltamethrin, *Aquatic Toxicology*, 33(3-4): 279-295.

Banaee, M., Sureda, A., Mirvaghefi, A.R. and Ahmadi, K. (2011). Effects of diazinon on biochemical parameters

- of blood in rainbow trout (*Oncorhynchus mykiss*), *Pesticide Biochemistry and Physiology*, 99(1): 1-6.
- Brander, S.M., Gabler, M.K., Fowler, N.L., Connon, R.E., Schlenk, D. (2016a). Pyrethroids pesticides as endocrine disruptors: molecular mechanisms in vertebrates with a focus on fishes. *Environ. Sci. Technol.*, 50: 8977–8992.
- Brander, S.M., Jeffries, K.M., Cole, B.J., DeCourten, B.M., White, J.W., Hasenbein, S., Fangue, N.A., Connon, R.E. (2016b). Transcriptomic changes underlie altered egg protein production and reduced fecundity in an estuarine model fish exposed to bifenthrin. *Aquat. Toxicol.*, 174: 247–260.
- Corcellas, C., Feo, M.L., Torres, J.P., Malm, O., Ocampo-Duque, W., Eljarrat, E., Barceló, D. (2012). Pyrethroids in human breast milk: occurrence and nursing daily intake estimation. *Environ. Int.*, 47: 17–22.
- Das, B.K. and Mukherjee, S.C. (2003). "Toxicity of cypermethrin in *Labeo rohita* fingerlings: biochemical, enzymatic and haematological consequences," *Comparative Biochemistry and Physiology C: Toxicology and Pharmacology*, 134(1): 109–121.
- El-Sayed, Y.S. and Saad, T.T. (2008). "Subacute intoxication of a deltamethrin-based preparation (Butox 5% EC) in monosex Nile tilapia, *Oreochromis niloticus* L," *Basic and Clinical Pharmacology and Toxicology*, 102(3): 293–299.
- Fırat, O., Cogun, H.Y., Yüzereroğlu, T.A. *et al.* (2011). "A comparative study of the effect of the pesticide (cypermethrin) and two metals (copper, lead) to serum biochemistry of Nile tilapia *Oeochromis niloticus*," *Fish Physiology and Biochemistry*, 37(3): 657–666.
- Kumar, A., Sharma, B. and Pandey, R.S. (2012). "Assessment of stress in effect to pyrethroid insecticides, λ-cyhalothrin and cypermethrin, in a freshwater fish, *Channa punctatus* (Bloch)," *Cellular & Molecular Biology*, 58(1): 153–159.
- Loteste, A., Scagnetti, J., Simoniello, M.F., Campana, M. and Parma, M.J. (2013). "Hepatic enzymes activity in the fish *Prochilodus lineatus* (Valenciennes, 1836) after sublethal cypermethrin exposure," *Bulletin of Environmental Contamination and Toxicology*, 90(5): 601–604.
- Nafiu, S.A., Zakariyya, M., Hassan, Z.Y. and Ahmad, M.K. (2020). Toxicological Effect of Inhaled Mosquito Incense Sticks Smoke on the Histology and Biochemical Responses in Experimental Rats. *Fudma*, *J. Sci.*, 4(4): 391-400.

- Sagar P., Singh S. and Singh A.P. (2017): Studies on Effect of Bifenthrin on Kidney Glycogen Content in *Channa Punctatus*. Annals of Natural Sciences, 3(3): 90-94.
- Sagar, P., Singh, S. and Singh, A.P. (2018). Effect of Bifenthrin Pesticide on Kidney Protein Content in *Channa Punctatus*. Asian journal of Agriculture & Life Sciences, 3(2): 55-58.
- Saha, S. and Kaviraj, A. (2009). "Effects of cypermethrin on some biochemical parameters and its amelioration through dietary supplementation of ascorbic acid in freshwater catfish *Heteropneustes fossilis*," *Chemosphere*, 74(9): 1254–1259.
- Ueyama, J., Kimata, A., Kamijima, M., Hamajima, N., Ito, Y., Suzuki, K., Inoue, T., Yamamoto, K., Takagi, K., Saito, I., Miyamoto, K.-I., Hasegawa, T., Kondo, T. (2009). Urinary excretion of 3-phenoxybenzoic acid in middle-aged and elderly general population of Japan. *Environ. Res.*, 109(2): 175–180.
- Ullah, S., Li, Z., Ul Arifeen, M.Z., Khan, S.U. and Fahad, S. (2019a). Multiple biomarkers based appraisal of deltamethrin induced toxicity in silver carp (*Hypophthalmichthys molitrix*). Chemosphere, 214: 519–533.
- Ullah, S., Zuberi, A., Alagawany, M., Farag, M.R., Dadar, M., Karthik, K., Tiwari, R., Dhama, K. and Iqbal, H.M.N. (2018). Cypermethrin induced toxicities in fish and adverse health outcomes: its prevention and control measure adaptation. *J. Environ. Manag.* 206: 863–871.
- Van den Berg, H., Zaim, M., Yadav, R.S., Soares, A., Ameneshewa, B., Mnzava, A., Hii, J., Dash, A.P., Ejov, M. (2012). Global trends in the use of insecticides to control vector-borne diseases. *Environ. Health Perspect.*, 120(4): 577–582.
- Velisek, J., Svobodova, Z., Packova, V. (2009). Effects of acute exposure of bifenthrin on some haematological, biochemical and histopathological parameters of rainbow trout (*Onchorhynchus mykiss*). *Veterin. Medic.*, 54(3): 131–137.
- Wang, X., Gao, X., He, B., Jin, Y.F.Z. (2017). Cis-bifenthrin causes immunotoxicity in murine macropalges. *Chemosphere*, 168:1375-1382.
- Wielgomas, B. and Piskunowicz, M. (2013). Biomonitoring of pyrethroid exposure among rural and urban populations in northern Poland. *Chemosphere*, 93(10): 2547–2553.

Cite this article-

Prem Sagar, Shalini Yadav, Shekhar Biswas, Neelam and Anand Pratap Singh, 2023, "Toxic effect of bifenthrin insectiside on the liver enzymes in *Channa punctatus* (Bloch.)" Journal of Science Innovations and Nature of Earth, Vol. 3(3), page-13-16

www.doi.org/10.59436/https://jsiane.com/archives3/3/94